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DATASHEET

Sucrose aCSF Instant Powder (packets)

Product overview

Name	Sucrose aCSF Instant Powder (packets)
Cat No	HB19127
Biological description	Sucrose artificial cerebrospinal fluid (sucrose - aCSF) is a widely used buffer as a protective cutting solution when making acute <i>ex-vivo</i> brain slices for electrophysiology experiments. This kit contains 10 instant powder packets. Simply dissolve the contents of each packet in dH ₂ O to a final volume of 1L, mix and bubble with carbogen to make 1L of sucrose aCSF at physiological pH.

Key features:

- Save time by using preformulated individual aCSF powder packets - each packet dissolves in seconds and there's no need to add Mg²⁺ or Ca²⁺
- More reproducible with each pack's highly accurate formulation - less error for better data.

Contains (in mM): Sucrose 65, NaCl 85, Glucose 10, NaHCO₃ 25, KCl 2.5, NaH₂PO₄ 1.25, CaCl₂ 0.5, MgCl₂ 7

Biological action	Buffer
Description	Preformulated instant powder packets to make sucrose artificial cerebrospinal fluid (sucrose - aCSF)

Solubility & Handling

Storage instructions	RT. Dissolve each pack in dH ₂ O to 1L final volume.
Handling	<p>Dissolve the contents of each packet in dH₂O to a final volume of 1000ml, mix well and bubble with carbogen (10-15 minutes) to make 1L of sucrose aCSF at physiological pH.</p> <p>Use immediately once opened.</p>
Important	This product is for RESEARCH USE ONLY and is not intended for therapeutic or diagnostic use. Not for human or veterinary use

Chemical Data

Kit contents	Preformulated packets. Each makes 1L of sucrose - aCSF.
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References

Acute brain slice methods for adult and aging animals: application of targeted patch clamp analysis and optogenetics.

Ting JT et al (2014) Methods in molecular biology (Clifton, N.J.) 1183

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[25023312](#)

Reduced long-term potentiation in hippocampal slices prepared using sucrose-based artificial cerebrospinal fluid.

Kuenzi FM et al (2000) Journal of neuroscience methods 100

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[11040373](#)

Repeated whole-cell patch-clamp recording from CA1 pyramidal cells in rodent hippocampal slices followed by axon initial segment labeling.

Oliveira LS et al (2021) STAR protocols 2

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[33644771](#)
